

## Research Article

## Drying Effect on Phytochemicals and Antioxidant Properties of Aqueous-Methanol Leave Extract of *Vernonia amygdalina* (Bitter Leaf)

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### Abstract

Traditional plants are known to possess nutraceutical properties. These properties are attributed to the types and amounts of phytochemicals present in them. *Vernonia amygdalina* (*V. amygdalina*) is widely distributed throughout tropical Africa. Research evidence has showed that *V. amygdalina* have been used traditionally to treat malaria, diabetes, diarrhea, hepatitis, skin disorders, cough, and wounds treatment. The present research focused on the effect of drying on phytochemical and antioxidant activities of aqueous-methanol leaf extracts of *V. amygdalina*. The plant samples were collected, washed and subjected to three difference treatments (sun-dried, shade-dried and fresh). The extracts were obtained using aqueous-methanol in ratio 1:2, then subjected to quantitative phytochemical screening and antioxidant, using reducing power assay (RPA), Total antioxidant capacity (TAC) and hydrogen peroxide inhibition ( $H_2O_2$ ) method. The results of the phytochemical screening shows that sun dried extract contain alkaloid ( $160.60 \pm 0.63 \mu\text{g/ml}$ ), phenol ( $281.40 \pm 3.57 \mu\text{g/ml}$ ), tannin ( $724.10 \pm 7.60$ ), saponin ( $0.18 \pm 0.00$ ) and flavonoid ( $24.22 \pm 1.10$ ). Shade dried results divulged total alkaloid ( $135.40 \pm 2.51$ ), Phenol ( $228.10 \pm 1.65$ ), tannin ( $168.00 \pm 0.57$ ), saponin ( $0.33 \pm 0.00$ ) and flavonoids ( $25.61 \pm 0.09$ ) whereas, fresh extract contained alkaloids ( $126.90 \pm 1.50$ ), Phenol ( $217.40 \pm 1.49$ ) Tannin ( $460.10 \pm 1.96$ ) saponin ( $0.36 \pm 0.00$ ) and flavonoids ( $4.70 \pm 2.44$ ). The highest concentration of Alkaloid, Phenols and tannin was obtained in sun dried while the highest flavonoids and saponin was obtained in shade dried and fresh extract respectively. The highest results of antioxidant were revealed in RPA ( $49.07 \pm 0.96$ ), TAC method ( $717.6 \pm 3.12$ ) at  $100 \mu\text{g/ml}$  while the  $H_2O_2$  highest activity was recorded for shade dried extract using the percentage inhibition method ( $64.02 \pm 0.43$ ) at  $1.0 \mu\text{g/ml}$ . The  $IC_{50}$  reveals that the sundried extract is more effective when compared with the rest as it has the lowest ( $0.677 \pm 0.007$ ) value of  $IC_{50}$ .

Keywords: Phytochemical, Antioxidant, *Vernonia amygdalina*.

## 1.0 INTRODUCTION

Plant medicine has been used by Most of the world's population due to its medicinal values and scarcity [1]. Medicinal plants

have been used to cure a wide range of illnesses since ancient times [2]. Despite recent technological

advancements in modern medicine, 75% of African communities still rely on traditional medicinal herbs for everyday healthcare. Many ailments can be treated using medicinal plants alone or in conjunction with other plants. Plants contain secondary metabolite chemicals, or non-nutritive compounds, called phytochemicals, which have the ability to prevent or treat disease. These nutrients are non-essential since the human body does not need them to survive. These compounds are produced by plants as self-defense, but new studies show that they can also serve as disease defense [3]. Bitter leaf, or *Vernonia amygdalina* Del., is a medicinal plant whose fresh leaves are very important to a human diet since they contain vitamin and mineral salts [4]. It is significant that *Vernonia amygdalina* has been utilized by humans for sustenance, maintenance, illness prevention, and treatment. Review of the literature has been discovered that the plant, particularly the leaf, is helpful in the ethnotherapy of diabetes [5], asthma [6], skin infections like ringworm, rashes, and eczema, schistosomiasis, malaria [7], measles, diarrhea, tuberculosis, abdominal pain, and intestine complaints, in addition to fevers, coughs, inducing fertility in women who are barren, and hyperlipidemia [8]. Properties of extracts have been used as an anthelmintic, antimalarial, anti-tumorigenic, bacteriostatic, and bactericidal action on some bacteria [9]. In particular, [5] documented the leaf extracts possess hypoglycemic and hypolipidemic effects. Several investigations have verified that *V. amygdalina* contains a variety of bioactive substances, including flavonoids, saponins, alkaloids, tannins, phenolics, terpenes,

steroidal glycosides, triterpenoids, and various forms of sesquiterpene lactones [10;11]. The aim of this study is to investigate the effect of drying on phytochemicals and antioxidant property of *Vernonia amygdalina* leaves.

## Materials and Methods

### Study Area

This study was carried out at the Department of Biochemistry, Bayero University Kano, Nigeria

### Sample Collection

The plant leaves were collected at school of continuous studies of Bayero University Kano, Kano State of Nigeria and identified at Department of Plant Biology Bayero University Kano with the following BUKHAN number 143 for future reference. The plant leaves were washed and divided into two parts for drying methods sun-dried, shade-dried and fresh leaves for 15days. The dried samples were pulverized into fine powder, while the fresh leaves were crushed, both the samples were weighed using electric weighing balance Elite 210-4, kilotech.

### Extraction of the plant leaves

Aqueous methanol extract of the powder sample was prepared by soaking 100 g of the sun dried, shade dried and fresh powdered leaves in 750 ml of absolute methanol and 1500 ml of distilled water (1:2) at room temperature for 72 hrs. The aqueous methanol extract was filtered and subsequently passed through cotton wool and concentrated using a rotary evaporator with the water bath set at 40°C to one-tenth its original volume and finally with a freeze

drier. The dried residue was then stored at 4°C. Portions of the extract was weighed and dissolved in distilled water for experimental analysis.

### Phytochemical Screening

#### Determination of Total Phenolic Content

The weight of each aqueous methanol extract sample was measured (0.01 g) and then they were dissolved in 10 ml of distilled water. 1 ml of the solution was moved into a test tube, then 0.5 ml of (2N) folin-ciocalteu reagent and 1.5 ml (20%) Na<sub>2</sub>CO<sub>3</sub> solutions were then added. The volume was raised to 8 ml with distilled water, vigorously shaken, left for 2 hours, and the absorbance was measured at 765 nm. The total phenolic content of the extract was determined by using a standard calibration curve with ascorbic acid solution and expressed in mg of ascorbic acid per gram of dry weight [12]. The phenolic content concentrations of the three extracts were determined using the ascorbic acid standard curve equation.

$$Y=0.0014x$$

Where y= absorbance

x=concentration

#### Determination of Total Flavonoids

Each leaf extract (0.01 g) weight was measured and then it was dissolved in methanol. After that, it was combined with 100 microliters of 20% aluminum trichloride and a drop of acetic acid before being thinned with methanol to 5 ml. An absorbance measurement at 415 nm was conducted following a 40-minute incubation period of the solution. A blank sample was created by

mixing 100 ml of plant extract with a drop of acetic acid, followed by dilution to 5ml with methanol. The standard rutin solution (0.5 mg/ml) in methanol was measured for absorbance under identical conditions [13]. The flavonoids content in three extracts was determined by using the rutin curve equation as per standard procedure.

$$Y=0.0062x+0.0012$$

#### Determination of Total Alkaloids

0.01 g of every sample was dissolved in 5 ml of 2N HCl and then filtered. 1 ml of the filtrate was moved into a separatory funnel and rinsed with 10 ml of chloroform. 1 ml of the solution was moved to a separatory funnel, followed by the addition of 5 ml of bromocresol solution and 5 ml phosphate buffer. The combination was agitated and the complex formed was fractioned with chloroform through vigorous shaking. The sample was placed in a 10 ml flask and mixed with chloroform until it reached the brim. The measurement of the complex's absorbance in chloroform was done at 470 nm [13]. Calculating the alkaloid concentrations of the three extracts was done using the standard aconitine curve equation.

$$Y=0.006x-0.003$$

#### Determination of Total Tannins

A 0.2 g sample was placed in a 50 ml plastic bottle, and then 20 ml of distilled water was added and shaken for an hour. The mixture was filtered in a 50 ml volumetric flask, topped up to the mark. 5 ml of the filtrate was pipetted into a test tube and mixed with 2 ml of 0.1M FeCl<sub>3</sub> in 0.1N HCl and 0.008 potassium ferrocyanide. The solution was left

to stand for 10 minutes before measuring the absorbance at 620 nm [13].

The concentration of the flavonoids contents of the three extracts were calculated following the standard gallic acid curve equation:

$$Y=0.0027x+0.0036$$

### Determination of Total Saponins Content

10 ml of distilled water was mixed with 0.1 g of each extract, followed by the addition of 10 ml of vanillin and 25 ml of 72% sulphuric acid to the solution. The mixture was thoroughly stirred and placed in a water bath at 60°C for 10 minutes, then cooled in ice water. The absorbance at 544 nm was measured and expressed as diosgenin equivalents (mg/g DE extracts) [14].

The concentration of the flavonoids contents of the three extracts were calculated following the standard diosgenin curve equation:

$$Y=5.636x$$

## ANTIOXIDANT ESTIMATION

### Determination of Antioxidant capacity by Phosphomolybdenum method

0.1mL of extract solution (100 µg/mL, 50 µg/mL, and 25 µg/mL) was combined with 1mL of reagent solution (0.6M sulphuric acid, 30mM sodium phosphate, and 4mM ammonium molybdate) to create a mixed aliquot. The tubes containing samples were closed and placed in a water bath at 95°C for 90 minutes to incubate. Following incubation, the reactant samples were allowed to cool down to room temperature,

then the absorbance of each aqueous solution was measured at 695 nm in comparison to a blank. A standard blank included 1 mL reagent solution and an equal amount of solvent as the samples, and then it was incubated under identical conditions as the other samples [15].

### Determination of Reducing power assay.

The aqueous methanolic leaf extract of *Vernonia amygdalina* was assessed for its reducing power capacity following the procedure outlined by Oyaizu [16]. The mixture had 1.0 mL of different concentrations of extracts (100 µg/mL, 50 µg/mL, and 25 µg/mL) and 2.5 mL of 0.2M sodium phosphate buffer. The blend was kept at a temperature of 50°C for half an hour, and the process was stopped by adding 2.5 mL of 10% trichloroacetic acid. After centrifuging at 3000r/min for 10 minutes, 2.5 mL of deionized water and 0.5 mL of 0.1% ferric chloride were included. The measurement of absorbance at 700 nm was done by comparing it to a blank solution consisting of deionized water and phosphate buffer. Ascorbic acid was employed as the standard. The findings were presented in terms of the amount of ascorbic acid equivalent per gram (AscAE/gm).

### Hydrogen Peroxide Scavenging (H<sub>2</sub>O<sub>2</sub>) Assay.

The ability of extracts to scavenge hydrogen peroxide was estimated by following the method of [17]. The extract was dissolved in a phosphate buffer at a concentration of 1 mg/ml. A quantity of 1 ml of extract was added to 3.4 ml of phosphate buffer (5mM, pH 7.4) followed by 600 µl of 400 mM H<sub>2</sub>O<sub>2</sub>.

The solution was kept at room temperature for 4 min and absorption was measured at 230 nm. Ascorbic acid was used as control. The percentage of hydrogen peroxide scavenging was calculated as follows:

$$\text{Scavenged H}_2\text{O}_2 (\%) = \frac{[A_i - A_t]}{A_i} \times 100$$

Where  $A_i$  was the absorbance of control and  $A_t$  was the absorbance of test samples.

## RESULTS AND DISCUSSION

### Result

#### Physical Properties of the Plant Extracts

Three extracts were obtained from *V. amygdalina* leaves. The aqueous methanol leaf extract of the fresh plant was brownish in colour and gummy in texture. The aqueous methanol leaf extract of the sun-dried plant was black-red in colour and sticky in texture. The aqueous methanol leaf extract of the shade dried plant was also greenish in colour and sticky in texture. Weight of plant sample before and after extractions were obtained as shown in Table 1

### Discussion

From the results obtained from the phytochemical quantitative determination of the sun-dried, shade dried and fresh aqueous methanol leaf extract of *V. amygdalina* reveals that Tannins was the highest concentration of all the phytochemical screened among the three samples (fresh, sun and shade dried) with a value of  $460.10 \pm 1.96$ ,  $724.10 \pm 7.60$  and  $168.00 \pm 0.57$  respectively. This is followed by Phenolic with corresponding values of  $217.40 \pm 1.49$ ,  $281.40 \pm 3.57$  and  $228.10 \pm 1.65$  for fresh, sun-

dried and shade-dried sample respectively. The least phytochemical present was saponins in all three sample (Table 2). The values of the phytochemicals are much higher in the Sun-dried samples when compared to the other two (Shade-dried and fresh) with a significant difference. This is then followed by shade-dried sample where phenolic, flavonoids, alkaloids and tannins were higher than those of fresh sample with values of  $168.00 \pm 0.57$ ,  $135.40 \pm 2.51$ ,  $25.61 \pm 0.09$  and  $228.10 \pm 1.65$  respectively. However, the Saponin was found to be highest in the Fresh sample ( $0.36 \pm 0.00$ ), then the shade-dried ( $0.33 \pm 0.00$ ) and the least value was for Sun-dried ( $0.18 \pm 0.00$ ). This result is in agreement with that of Misari [18] where, from his results, the Saponin level was found to be least while phenolic; flavonoid and tannin were the highest. In another work by [19], the Sun-dried sample contains higher amount of all the five phytochemicals screened as compared with the shade-dried. However, their result does not include values for fresh sample. From the results obtained from the Anti-oxidant property using the Reducing power assay (Table 3), it was observed for all the three samples, the values increases with an increase in the concentration of the extract used with 100  $\mu\text{g/ml}$  having the highest values, then 50  $\mu\text{g/ml}$  and the least values were recorded for plant extracts at 25  $\mu\text{g/ml}$ . Comparing the three extract samples it was found out that the values obtained for Shade-dried samples were the highest ( $49.07 \pm 0.96$ ,  $41.98 \pm 0.65$  and  $32.66 \pm 0.83$ ) for the three (3) different concentrations respectively (Table 2). This is then followed by those of Sun-dried plant extracts and the least was recorded for fresh

sample Mbakwem [20] also observed similar result from his study. He recorded that the Anti-oxidant property of *V.amygdalina* leaf extract using reducing power assay is concentration dependent as the highest values were recorded at a concentration of 250  $\mu\text{g/ml}$  and the least at 50  $\mu\text{g/ml}$ . Table 4 shows the result for the Phosphomolybdate assay (total anti-oxidant capacity) of the three plant extracts at different concentrations of 100  $\mu\text{g/ml}$ , 50 $\mu\text{g/ml}$  and 25 $\mu\text{g/ml}$ . From the table, the highest activity was recorded for fresh plant extract at 100  $\mu\text{g/ml}$  (717.6 $\pm$ 3.12), followed by sun-dried extract at 50  $\mu\text{g/ml}$  (703.7 $\pm$ 9.45), sun-shade at 25  $\mu\text{g/ml}$  (695.9 $\pm$ 20.4). This result shows that the Phosphomolybdate activity of three extract of *V.amygdalina* is not concentration dependent. Table 5: shows the result for hydrogen peroxide %inhibition assay of the three plant extracts. The value for shade-dried samples increased steadily as the concentration increases from 14.60% (0.2mg/ml) to 54.02% (1.0mg/ml). The value of sun dried rise gradually and slowly as the concentration is increased from 0.2 mg/ml till 0.6mg/ml from where the curve continues to rise sharply to attain a peak value of 55.45% at 1.0mg/ml. The fresh sample curve shows a very rapid increase as the concentration of extract increased to obtain a peak value of 67.03% at the concentration of 0.8 mg/ml and then began to fall as the concentration increased to 1.0 mg/ml. The result show that all the three extracts have a very good ability as free radical scavenger and their ability is concentration dependence i.e. increased as the concentration increases. Comparison of the three extract (sun dried, shade dried and

fresh sample) show that the highest peak was observed for fresh sample (67.03%) at 0.8mg/ml and the least from shade dried (14.60%) at 0.2 mg/ml. In Table 6, among the three extracts subjected to reducing power assay test in the study, it was observed that the fresh extract of *V. amygdalina* leaves have the highest concentration with  $\text{IC}_{50}$  of 166.971  $\mu\text{g/ml}$  This is consecutively followed by Sun dried extract with  $\text{IC}_{50}$  of 138.535  $\mu\text{g/ml}$  and shade dried extract give the lowest concentration of  $\text{IC}_{50}$  of 83.905  $\mu\text{g/ml}$ . Hence comparing the sun-dried extract and fresh extract has less effective compared to shade-dried. On hydrogen peroxide inhibition percentage, the three extract show lowest concentration with  $\text{IC}_{50}$  on sun-dried (0.677  $\mu\text{g/ml}$ ) extract followed by shade dried extract with  $\text{IC}_{50}$  (1.848  $\mu\text{g/ml}$ ) and highest in fresh extract with  $\text{IC}_{50}$  (3.084  $\mu\text{g/ml}$ ). And on the phosphomolybdate assay shade dried extract record highest activities (143.791  $\mu\text{g/ml}$ ) followed by fresh extract (5.440  $\mu\text{g/ml}$ ) and sun dried extract was the least (5.401  $\mu\text{g/ml}$ ). For the combined  $\text{IC}_{50}$  of the three tests ( $\text{H}_2\text{O}_2$ , RPA and TAC) for the antioxidant property of *V. amygdalina*, it was observed that the sun-dried extract show least combined  $\text{IC}_{50}$  value of (0.677  $\pm$  0.007) for  $\text{H}_2\text{O}_2$  and (5.401  $\pm$  0.086) for TAC. Lower value of  $\text{IC}_{50}$  indicates a higher antioxidant property hence sun-dried extract is more effective compared to the shade and fresh samples. However, shade-dried samples had higher activity than the fresh samples with an overall highest  $\text{IC}_{50}$  value for the three (3) tests conducted.

**Table 1: Physical Properties of Leaf Extracts from *V. amygdalina***

Property	FRESH	SUN-DRIED	SHADE-DRIED
Weight of plant sample (g)	100.237	100.680	100.003
Weight of plant extract (g)	21.098	19.641	14.138
Colour of extract	Brownish	Greenish-Black	Greenish
Texture of extract	Gummy	Sticky	Sticky

**Table 2 Quantitative Phytochemical Analysis**

Phytochemicals	Fresh	Sun-dried	Shade-dried
Tannins ( $\mu\text{g/ml}$ )	460.10 $\pm$ 1.96 <sup>a</sup>	724.10 $\pm$ 7.60 <sup>b</sup>	168.00 $\pm$ 0.57 <sup>c</sup>
Saponins ( $\mu\text{g/ml}$ )	0.36 $\pm$ 0.00 <sup>a</sup>	0.18 $\pm$ 0.00 <sup>b</sup>	0.33 $\pm$ 0.00 <sup>c</sup>
Alkaloids ( $\mu\text{g/ml}$ )	126.90 $\pm$ 1.50 <sup>a</sup>	160.60 $\pm$ 0.63 <sup>c</sup>	135.40 $\pm$ 2.51 <sup>a</sup>
Flavonoids ( $\mu\text{g/ml}$ )	4.70 $\pm$ 2.44 <sup>b</sup>	24.22 $\pm$ 1.10 <sup>a</sup>	25.61 $\pm$ 0.09 <sup>a</sup>
Phenolic ( $\mu\text{g/ml}$ )	217.40 $\pm$ 1.49 <sup>a</sup>	281.40 $\pm$ 3.57 <sup>b</sup>	228.10 $\pm$ 1.65 <sup>a</sup>

Values are mean  $\pm$  SD (n=3). Value with same superscript on the same row shows no significant difference at  $p > 0.05$ .

**Table 3: Reducing Power Assay of Shade-Dried, Sun-Dried and Fresh Aqueous Methanol Leaf Extract of *V. amygdalina***

Samples	Extract Concentration ( $\mu\text{g/ml}$ )		
	100 $\mu\text{g/ml}$	50 $\mu\text{g/ml}$	25 $\mu\text{g/ml}$
Sun-dried	27.85 $\pm$ 1.16 <sup>a,*</sup>	25.82 $\pm$ 1.79 <sup>a,*</sup>	26.84 $\pm$ 1.13 <sup>a,*</sup>
Shade-dried	49.07 $\pm$ 0.96 <sup>a,***</sup>	41.98 $\pm$ 0.65 <sup>b,***</sup>	32.66 $\pm$ 0.83 <sup>c,***</sup>
Fresh	22.10 $\pm$ 1.077 <sup>a,**</sup>	21.90 $\pm$ 1.13 <sup>a,**</sup>	21.65 $\pm$ 1.10 <sup>a,**</sup>

Values are mean  $\pm$  SD (n=3). Value with same superscript (a, b, c) on the same row shows no significant difference at  $p > 0.05$ . Values with same superscript (\*, \*\*, \*\*\*) down the column shows no statistical significant at  $p > 0.05$ .

**Table 4: Phosphomolybdate Assay extracts Concentration ( $\mu\text{g/ml}$ )**

Samples	Extract concentration ( $\mu\text{g/ml}$ )		
	100 $\mu\text{g/ml}$	50 $\mu\text{g/ml}$	25 $\mu\text{g/ml}$
Sun	689.5 $\pm$ 14.5 <sup>a,*</sup>	703.7 $\pm$ 9.45 <sup>b,*</sup>	695.9 $\pm$ 20.4 <sup>ab,*</sup>
Shade	27.26 $\pm$ 3.06 <sup>b,**</sup>	24.29 $\pm$ 0.43 <sup>a,**</sup>	25.39 $\pm$ 0.16 <sup>a,**</sup>
Fresh	717.6 $\pm$ 3.12 <sup>a,***</sup>	649.4 $\pm$ 37.7 <sup>b,***</sup>	656.5 $\pm$ 90.7 <sup>c,***</sup>

Values are mean  $\pm$  SD (n=3). Value with same superscript (a, b, c) on the same row shows no significant difference at  $p > 0.05$ . Values with same superscript (\*, \*\*, \*\*\*) down the column shows no statistical significant at  $p > 0.05$ .

**Table 5:** Hydrogen Peroxide Inhibition% Extracts Concentration (mg/ml)

Sample	Extract concentration (mg/ml)				
	0.2	0.4	0.6	0.8	1.0
Sun	35.04 $\pm$ 13.70 <sup>a,*</sup>	37.26 $\pm$ 8.10 <sup>a,*</sup>	39.03 $\pm$ 0.27 <sup>b,*</sup>	52.21 $\pm$ 4.08 <sup>c,*</sup>	55.95 $\pm$ 0.96 <sup>d,*</sup>
Shade	14.60 $\pm$ 11.92 <sup>a,**</sup>	41.08 $\pm$ 6.42 <sup>b,**</sup>	54.50 $\pm$ 2.63 <sup>c,**</sup>	54.71 $\pm$ 2.45 <sup>c,**</sup>	64.02 $\pm$ 0.43 <sup>d,**</sup>
Fresh	28.90 $\pm$ 8.49 <sup>a,***</sup>	45.81 $\pm$ 5.79 <sup>b,***</sup>	57.03 $\pm$ 5.96 <sup>c,***</sup>	67.03 $\pm$ 1.29 <sup>d,***</sup>	53.43 $\pm$ 2.10 <sup>c,**</sup>

Values are mean  $\pm$  SD (n=3). Value with same superscript (a, b, c, d) on the same row shows no significant difference at  $p < 0.05$ . Values with same superscript (\*, \*\*, \*\*\*) down the column shows no statistical significance at  $p < 0.05$ .

The result for the concentration of the extracts producing 50% Reducing Power Assay, Hydrogen Peroxide Inhibition% and Phosphomolybdate Assay (IC<sub>50</sub>) are shown in the Table 6 below.

**Table 6: Concentration of Extracts at 50% Activity for Sun dry, Shade dry and Fresh Extracts of *V. amygdalina* leaves.**

Extracts	H <sub>2</sub> O <sub>2</sub>	RPA	TAC
Sun-dried	0.677 $\pm$ 0.007 <sup>a,*</sup>	138.535 $\pm$ 3.717 <sup>b,**</sup>	5.401 $\pm$ 0.086 <sup>c,**</sup>
Shade-dried	1.848 $\pm$ 0.097 <sup>a,**</sup>	83.905 $\pm$ 0.978 <sup>b,*</sup>	143.791 $\pm$ 8.831 <sup>c,***</sup>
Fresh sample	3.084 $\pm$ 0.696 <sup>a,***</sup>	166.971 $\pm$ 7.605 <sup>b,***</sup>	5.440 $\pm$ 0.038 <sup>a,**</sup>

Values are mean of 3 determination  $\pm$  Standard deviation. Values with the same superscript (a, b, c) across the same row show no significantly different with each other at  $P < 0.05$ .

Values with different superscript (\*, \*\*, \*\*\*) down the group show significantly different with each other at  $P < 0.05$ .

**H<sub>2</sub>O<sub>2</sub>**= hydrogen peroxide inhibition%

**RPA**= reducing power assay

**TAC**= phosphomolybdate assay

## CONCLUSION

It was concluded that, drying process has a profound effect on some of the phytochemicals of the leave extract of *V. amygdalina*. Exposure to the sun-dried extract increases the amount of alkaloids, phenols and flavonoids, tannin compared to the fresh sample extract while saponnin

decreases in the sun-dried extract. Shade-dried extract showed increase in the alkaloids, phenol and flavonoids content compared to the fresh sample extract while tannin and saponnin content decreases in the shade.



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